13 Role of Arbuscular Mycorrhiza and Associated Microorganisms in Phytoremediation of Heavy Metal-Polluted Sites

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CONTENTS

13.1 INTRODUCTION

Various technologies exist that enable the detoxification/deactivation and removal of toxic compounds from the soil, mostly based on physicochemical extraction methods. They are costly and totally destroy soil microorganisms. The restitution of life on polluted sites or areas that were subjected to conventional technologies usually takes a very long time, is difficult, and often requires human intervention.

Phytoremediation is an alternative to physicochemical methods; it involves the use of plants in the process of decreasing the level of toxic compounds in soil, stabilizing the soil, and inhibiting erosion [1]. However, plants need appropriate below-ground ecosystems to establish diverse communities, especially on difficult sites [2–4]. Mycorrhizal fungi play a key role in increasing the volume of soil explored by the plant in search for nutrients and trace elements. Their activity directly or indirectly influences the microbial populations, qualitatively and quantitatively, including bacteria and fungi from the zone called the mycorrhizosphere.

The present chapter will focus on arbuscular mycorrhizal fungi (AMF). Under natural conditions, they are accompanied by bacteria such as legume symbiotic nodular bacteria, plant growth-promoting rhizobacteria (PGPR), and fungi, including other mycorrhizal symbionts and saprobic fungi. All of these organisms build specific consortia that influence the plant by means of interactions with abiotic [5] and biotic components of the soil [6] or stimulate plant growth through the production of vitamins and hormones [7,8]. Rebuilding/establishing such consortia is of utmost importance for the effectiveness of phytoremediation processes.

13.2 MYCORRHIZA AND ITS ROLE IN THE ENVIRONMENT

Mycorrhizal fungi are widespread and form symbiosis with a large majority of plant species on Earth [2]. A common trait of all mycorrhizal fungi is that their mycelium overgrows the soil surrounding the plant roots; the hyphal net stabilizes the soil and furthermore produces substances that bind or glue soil particles together [9]. The way in which the root and its surface are colonized depends on the type of mycorrhiza.

There are two main types of mycorrhiza: ecto- and endomycorrhiza. In the first case, the mycelium forms a more or less compacted fungal mantle on the surface of the root. Its protective properties depend on the species of the symbiotic fungus, the mantle's water-absorbing capacity, the production of pigments, and organic acids. The mycelium penetrates between cortical cells of the root, forming the so-called Hartig net, which is the site of exchange of compounds between the partners. Ectomycorrhiza is formed by several thousands of fungal species, more or less specific towards host plant species (usually trees from the temperate zone). These trees are obligate symbionts.

Endomycorrhiza is far more diverse. Its characteristic feature is the possibility to penetrate not only spaces between cells, but also the inside of live cortical cells, crossing the cell wall and then developing in touch with the plasma membrane of the plant cell. This type of symbiosis includes orchid, ericoid, and arbuscular mycorrhiza. In the first two types, the mycelium forms coils inside cortical cells, but in arbuscular mycorrhiza, characteristic tree-like structures termed arbuscules develop. Arbuscular mycorrhiza (AM) is the most widespread type of mycorrhiza, occurring in 80% of plant species; it is formed by about 120 species of fungi belonging to the Glomeromycota [10]. This symbiosis is believed to be phylogenetically the most ancient type of mycorrhiza.

Molecular and paleobotanical studies seem to support the hypothesis of a close relationship of the AMF with plants since they appeared on land [11,12]. This mycorrhiza plays a key role in the productivity, stability, and diversity of natural ecosystems. Natural soils with low levels or completely devoid of AMF propagules are rare. Several factors can influence the quantity (i.e., the number of propagules) and the quality (i.e., the composition in species) of AM fungi in the soil. The presence of heavy metals and/or other pollutants, the use of amendments to remediate pollution, and the kind of vegetation heavily affect the composition and abundance of the Glomalean fungi [13–15].

The disappearance of the propagules leads to serious consequences, such as the degradation of plant communities; decreased availability of essential elements; and loss of ecosystem stability. Among examples in which it is necessary to introduce AMF propagules during creation and rebuilding of plant communities are sites resulting from volcanic activity and cutting down of forests; industrial wastes; postmining open areas; excessively fertilized agricultural lands; and soils strongly polluted by toxic compounds such as heavy metals (HM) and xenobiotics [2]. In such situations, the introduction of mycorrhizal inoculum involving selected fungal strains adapted to survive in a given toxic environment and under given climatic conditions becomes a key tool in decreasing the toxicity of these compounds to the plants and in establishing a stable vegetation cover.

Fungi adapted to polluted soils should be a choice of preference for the production of inoculum for soil remediation. The number of spores in polluted areas can be affected by the presence of heavy metals, but different fungi show different sensitivity [15] and species-specific (or even strainspecific) behaviors can be observed. In order to reduce the costs of inoculation, the choice of plants is also a relevant point. Plants should be efficient in removing or stabilizing the pollutants, and they should also promote the establishing of strong mycorrhizal and microbial communities because different plant species can affect the species composition of the Glomalean community [15].

13.3 PHYTOREMEDIATION AND THE BEGINNING OF INTEREST IN MYCORRHIZA

At first, the necessity to include soil microorganisms in phytoremediation was neglected. People used compounds that increase the availability of toxic compounds, therefore stimulating the accumulation of metals in plants [16–18], as well as fertilizers to boost plant biomass production [19]. The most efficient varieties were selected; techniques involving genetic engineering were also used [20–23]. The plants' ability to produce organic compounds influencing the rhizosphere and increasing the availability of metals was also acknowledged [24–26].

Among plants that produce high amounts of organic acids in the rhizosphere, researchers' attention was drawn to the order *Lupinus*. Its cultivation can successfully replace the use of chemicals increasing the availability of soil metals. At last the fact that the activity of microorganisms is a factor strongly influencing the processes of mobilizing and immobilizing metals, by means of precipitating sulphides and hydrated iron oxides or their binding to polysaccharides, was noticed [24,27]. Elements such as Pb, Zn, and Cu can also bind to carbonates and oxalates produced by microorganisms [28]. Metals can as well bind to functional groups localized on the surface of the microorganisms' cell walls [29].

Biological methods of cleaning up contamination mainly use bacteria and saprobic fungi; the role of mycorrhizal fungi is still underestimated. Well-developed mycorrhiza can enhance plant survival in difficult areas because it increases the availability of biogens; reduces stress due to low water availability; increases the resistance to pathogens; stimulates the production of phytohormones; and generally improves the soil structure. These factors can significantly enhance bioremediation.

Among the usually considered bioremediation practices, special attention is due to phytostabilization, phytodegradation, and phytoextraction. Briefly, phytostabilization involves the immobilization of toxic compounds in the soil by means of plants that reduce soil erosion; leaking of contaminants into the ground waters; and their dispersion through wind erosion [30]. Phytodegradation includes various metabolic processes of plants and accompanying microorganisms leading to the breaking up of organic compounds such as polyaromatic hydrocarbons, pesticides, and explosives. Phytoextraction takes advantage of the ability of plants to hyperaccumulate metals. Plants are considered useful if they can take up over 1% of a given metal in the dry mass of their shoots. Such plants are grown on the given area and their above-ground parts are harvested, dried, and burned [31,32].

According to Gleba et al. [33], during phytoextraction the "giant underground networks formed by the roots of living plants function as solar driven pumps that extract and concentrate essential elements and compounds from soil and water." This observation has some important implications and consequences.

First, because heavy metals are taken up and transported in water solution, increased plant transpiration would increase metal translocation to the shoot. There is no doubt that mycorrhizal colonization affects the water relations of plants (Smith and Read [2] and references therein). A number of papers indicate that transpiration rates of mycorrhizal plants are significantly higher than those observed in nonmycorrhizal ones [34–39]. Mycorrhizal root systems are usually more branched [40] and therefore they present a larger absorbing surface even in the absence of changes in root biomass [34]. Also, the increased leaf area can be an important factor leading to increased transcription [41]; however, even comparing plants of the same size and root system length, the transpiration rates of mycorrhizal plants remain superior, due to the reduced stomatal resistance [38].

It has been noted that the high stomatal resistance observed in P-deficient nonmycorrhizal plants is a nutritional effect [38,39]. Nevertheless, stomatal behavior is affected by hormonal changes, which can depend on P nutrition as well as on mycorrhizal colonization [35,42–44]. Abscissic acid (ABA) is known to block transpiration and, consequently, metal accumulation in shoots [17]. According to Allen [35], ABA concentrations in leaves of *Bouteloua gracilis* decreased following mycorrhizal colonization by *Glomus fasciculatum*; on the other hand, Danneberg et al.

[43] report higher concentrations of ABA in leaves and roots of maize colonized by *Glomus* sp. (isolate T6) in comparison to nonmycorrhizal plants. Once more, the different combination of plant and fungus species might be important, as well as the growth conditions and the methods utilized for the measurements.

In the second place, roots and hyphae of mycorrhizal root systems explore an incredibly larger volume of soil in comparison with nonmycorrhizal root systems. Even if the contribution of the external hyphae to water uptake and its translocation to the roots has not been clarified [2], it was proved that the external mycelium may contribute to the uptake and translocation of some heavy metals (including Zn, Cu, and Cd [45–47]) to the host roots.

The preceding considerations show, once more, how important it is to gain deeper knowledge on the basic functioning of the mycorrhizal symbiosis for its exploitation in biotechnology and environmental applications.

13.4 MYCORRHIZA IN PHYTOSTABILIZATION

Mycorrhiza proved to be especially useful in phytostabilization. Although the first plants that colonize areas with increased heavy metal levels usually belong to nonmycorrhizal species [48,49], the development of a dense vegetation cover and improvement of the soil structure require the presence of symbiotic fungi. This is especially important for sites where postflotation material originating from zinc and lead ore processing is deposited. Such material is almost devoid of nitrogen and phosphorus compounds, has poor water-holding capacities, and is vulnerable to wind erosion [50]. Possible mechanisms of improved resistance of AMF-colonized plants to HMs include the enhancement of nutrient uptake, particularly phosphorus, and water supply [51,52], as well as metal sequestration through the production of binding substances or absorption of metals by microbial cells [47,53].

Recently, a gene called *GmarMT1,* encoding for a fungal metallothionein, has been identified in *Gigaspora margarita* (BEG 34) [54]. Metallothioneins (MT) are Cys-rich polypeptides able to chelate metal ions and important in the buffering of their intracellular concentration. Heterologous complementation in yeast revealed that the polypeptide encoded by *GmarMT1* confers increased tolerance to Cu and Cd. The gene expression in the symbiotic mycelia is up-regulated upon Cu exposure [54]. Spontaneous colonization of polluted substrate by arbuscular fungi takes a long time. However, it is possible to introduce propagules of selected strains of mycorrhizal fungi in the form of inoculum. Individual strains show a pronounced diversity in the effectiveness of metal binding and therefore also in reducing the toxicity of the substratum. Because the mycelium of certain strains of species, like *Glomus mosseae*, are tolerant to high heavy metal concentrations, they can bind a few times more metals than the mycelium of a saprobic species commonly used in bioremediation — *Rhizopus arrhizus* [55].

It was demonstrated for cadmium and zinc that, although these elements are detected in the mycelium developing inside plant roots, their accumulation in above-ground parts might be limited [46,56]. However, the analysis of tissue concentration and total shoot uptake of Cd, Zn, and Pb in *Plantago lanceolata*, grown in rhizoboxes on substratum collected from zinc wastes, inoculated with a number of arbuscular mycorrhizal fungal (AMF) strains has shown that metal uptake by the plant differs depending on AMF strain/species [57] [\(Figure 13.1\).](#page-4-0)

The ability to bind and detoxify heavy metals in underground parts of plants might be of importance also for the stimulation of the growth of crops cultivated on polluted soils. Such plants were inoculated with a *Glomus intraradices* strain isolated from a metallophyte — *Viola calaminaria* [56,58]. AMF also eliminated the toxic effect of Cd on several pea genotypes [59]. It was noted that strains isolated from polluted areas are far more useful than strains originating from nonpolluted sites [60–62]. Differences in the effectiveness of metal detoxification and accumulation exist also among strains and species occurring on polluted sites [63]. This underlines the importance of the identification

FIGURE 13.1 Pb content in shoots of three maize varieties, mycorrhizal/nonmycorrhizal and with/without EDTA treatment, as examples of different behavior patterns. Different letters above bars indicate statistically significant differences at *p* < 0.05. (Modified from Jurkiewicz, A. et al., *Acta Biol. Cracov. Bot*., 46, 2004.)

and characterization of the strains, aimed at the selection of the most effective ones. The selected strains should also effectively compete with other fungi that might occur on the given site.

The inoculation of agricultural fields with mycorrhizal fungi in nonpolluted areas often does not improve the situation due to the presence of native strains, which are much better adapted to the given soil conditions, and therefore seems to be ineffective; however, in the case of polluted places that suffer from reduced propagule number and decreased range of fungal species, inoculation is far more effective. The recently developed molecular methods enable scientists to track the fate of the introduced strain in pot cultures as well as in field-collected material. Specific primers exist for a range of species and strains and allow detection of the presence of the introduced fungi in root samples stained to reveal the intraradical mycelium, using the nested PCR method [63–65]. In addition to analysis of the parameters of the fungal partner, it is also necessary to investigate the mechanisms that enable the plant to tolerate metals transferred from the mycelium. Plants show a range of reactions to heavy metals [66]; they can also regulate the degree of mycorrhizal colonization [67].

When selecting plant species for phytostabilization, special attention is usually given to grasses. Although C3 grasses are, under natural conditions, medium or poorly mycorrhizal, they become strongly colonized by AMF on industrial wastes [68–71] and in areas seriously polluted by heavy metals [72–74]. Introduction of the AMF inoculum simultaneously with mixtures of grasses adapted to given conditions is important for the grasses and may be the source of propagules for the establishment of trees such as *Acer* and *Populus* [75]. Similarly, the establishment of ectomycorrhizal tree plantations such as pine or birch (a common practice on industrial wastes) allows improvement of the structure of the soil and increases the soil organic matter content; this in turn creates better growth conditions for herbaceous plants and their symbiotic organisms [76].

Similarly to arbuscular mycorrhizae, ectomycorrhizae stimulate the growth of trees, protect them against pathogens, and may alleviate soil toxicity [77]. Ectomycorrhizal fungi appear on heavily polluted sites faster than arbuscular ones. This is because they form small spores in fruitbodies that usually expose the reproductive layer above the ground level, enabling easy, longdistance dispersion by wind. Individual species and strains of ectomycorrhizal fungi also show diversity in the effectiveness of protecting trees on polluted soils [53,78,79]. This phenomenon is clearly visible when comparing strains isolated from polluted and nonpolluted sites in laboratory conditions [80]. The mycelium can immobilize heavy metals and reduce their transfer to plant tissues, which is regarded as an important protection mechanism allowing trees to survive in polluted areas [53,74].

The phenomenon is explained by the binding of elements by pigments deposited on the surface of extraradical mycelium [60,81], within the hyphal wall [82,83], and in phosphate-rich vacuolar granules [81]. The biofiltration effect of the extraradical mycelium is the most pronounced. The fungal mantle can sometimes also play this role. Such a situation was described in *Rhizopogon roseolus* and *Suillus luteus* from zinc wastes in southern Poland [84,85]. In mycorrhizae of pines with the mentioned fungal species, a typical gradient of heavy metals, decreasing towards the inside of the mantle, was observed. The selection of mycorrhizal strains for the inoculation of tree seedlings planted in polluted areas is important for their establishment.

13.5 MYCORRHIZA IN PHYTODEGRADATION AND PHYTOEXTRACTION

Phytodegradation consists of accelerated degradation of polluting organic compounds, such as hydrocarbons, pesticides, or explosives, in the presence of plants. Existing technologies involve mostly saprobic bacteria and fungi [86–88]. Plants with an abundant root system also have a favorable effect on the degradation of polyaromatic hydrocarbons (PAH) [89,90]. The introduction of rhizosphere microorganisms into such cultures is an alternative to chemical compounds that increase the availability of toxic substances [89,91].

Arbuscular and ectomycorrhizal fungi can enhance phytodegradation [92]. Although the number of propagules of arbuscular fungi decreases with increasing concentration of xenobiotics [93], they can still stimulate plant growth by decreasing the stress related to low phosphorus availability [94] and water deficiency [95]; they can also boost the production of oxidation enzymes [96]. Ectomycorrhizal fungi can additionally produce enzymes taking part in preliminary or intermediate stages of xenobiotics' decomposition [52,97], which enables their further decomposition by other rhizosphere organisms [98–100]. In addition, soil polluted with organic compounds is usually rich in heavy metals. Although these metals cannot be degraded, development of the mycorrhizal mycelium can efficiently alter their availability and plant growth conditions.

Recently, attention has been paid to *Phragmites australis*, which is commonly used for constructed wetlands designed to treat organic effluents [101–103]. *P. australis* was so far believed to be nonmycorrhizal, but again it was recently found to form the symbiosis with enhanced frequency when the water level was reduced and during the flowering time [104]. The presence of potentially mycorrhizal plants in constructed wetlands might be important to enhance phytostabilization and improve the restoration of biodiversity in areas where the processes had ceased.

The least attention was paid to the use of mycorrhiza in phytoextraction. Recent years have brought an increase of interest in the hyperaccumulation of heavy metals by plants, due to the commercial potential of phytoremediation in cleaning up contaminated soil [19,105,106], and as a method to mine metals from low-grade ore bodies [107–110]. Although several reports have been issued on arbuscular mycorrhiza of plants occurring on heavy metal-rich soils such as serpentines [111,112] or strip mines [70,76], arbuscular mycorrhiza was only recently reported in a few hyperaccumulating species belonging to the Asteraceae family growing on nickel-enriched ultramafic soils in South Africa. All plants were found to be consistently colonized by AM fungi, including an abundant formation of arbuscules.

Among them, the most important for phytomining is *Berkheya coddii*, which is capable of accumulating up to 3.8% of Ni in dry biomass of leaves under natural conditions and produces a high yield exceeding that for most hyperaccumulators [113]. The species can also provide an excellent model for laboratory studies on mechanisms mediated by AMF fungi that allow for the phytoextraction process. It has been also shown to form well-developed mycorrhiza under greenhouse conditions. Preliminary results have shown that *B. coddii* inoculated with native fungi (*Gigaspora* sp. and *Glomus tenue*) had not only higher shoot biomass but also significantly increased Ni content (over two times) in comparison to noninoculated plants. This finding greatly contrasts

with the conventional opinion that the presence of AMF reduces the uptake of trace elements if they occur in excessive amounts [53].

Mycorrhizal colonization was also reported in Zn and Pb hyperaccumulating *Thlaspi praecox* from the Alps; still, the colonization level of this plant is rather low and decreases with increasing content of heavy metals in the soil [114]. In addition, it is thus far not possible to obtain the formation of mycorrhiza of this group of plants under laboratory conditions; this makes the interpretation of field data hard to confirm.

Although mycorrhiza does not necessarily stimulate phytoextraction, its potential to increase the biomass of the plants, improve soil conditions, and protect the plants from pathogens offers important reasons to include this phenomenon in further research. Three possibilities to increase phytoextraction are being proposed: (1) transgenic plants; (2) hyperaccumulators or high biomass producing crops such as maize, especially for soils relatively less polluted [115,116], treated with chemical chelating substances such as EDTA or sulphur; and (3) stimulating the development of or introducing rhizosphere organisms that will increase the uptake of metals by the plants.

The biotechnological approach aims at producing genetically modified plants characterized by increased tolerance to toxic compounds, higher biomass, and high uptake of heavy metals. A number of transgenic plants have already been obtained by transferring appropriate genes from bacteria or yeasts (see, for example, references 117 through 119) or by generated somatic hybridization between plants such as *Brassica napus* and *Thlaspi caerulescens* [120]. Most transgenic plants have, thus far, only been tested under artificial conditions [121] and they still need further research before the application phase will start. Also, the transformation of AM fungi has been approached. The identification of genes with similar functions can be very important for understanding of the mechanisms of resistance and tolerance to heavy metals and for selection of the fungal strains most suitable for phytostabilization and phytoremediation.

However, although the transformation of many animal, plant, and fungal (e.g., *Saccharomyces*) species is now a relatively easy practice, standard protocols for the transformation of AM fungi are not available yet [122]. Beyond technical problems, the huge diversity of the fungal genome inside one single isolate [123] and the lack of knowledge about the factors controlling the expression of this diversity represent a major problem for an effective exploitation of this kind of approach. In addition, the below-ground environment in which the fungi live and their vegetative reproduction make control of the spread of the transformed fungi very difficult, suggesting a very cautious and careful introduction.

The use of synthetic chelates has been proposed because the amount of metals extracted from the soil by plants depends largely on the availability of the metals. In most soils, even highly polluted ones, only a relatively small percentage of the total metal pool is available to plants. These compounds mobilize metal ions and displace them into the soil solution. Among a variety of chelates tested by Huang et al. [116], EDTA was demonstrated to be the most effective in mobilizing Pb [124], showing that it also increased the availability of other metals such as Cd, Cu, Ni, and Zn.

Experiments carried out on maize show that this common crop can take up as much as 3000 mg kg⁻¹ Pb in shoots when grown in laboratory conditions with EDTA (0.5 g kg⁻¹ of soil) [116]. A study carried out on 15 commercially available Polish maize varieties inoculated with an AMF strain and treated with EDTA showed that most maize varieties cultivated on metal-rich substratum had higher shoot biomass; this clearly confirmed the role of mycorrhizal fungi in phytoremediation practices, although large differences between varieties have been observed [125].

The data show a large diversity in the effectiveness of phytoextraction among different varieties of the same species. This finding stresses the necessity to screen a large number of cultivars in order to select the best ones for further use. Moreover, the effect of individual varieties might vary when chelators such as EDTA are used [\(Figure 13.2\).](#page-7-1) Although the use of chemical amendments should be considered carefully, heavy metal release into water flowing out from EDTA-treated pots was found to be substantially lower in the case of mycorrhizal plants than in nonmycorrhizal plants.

FIGURE 13.2 Heavy metal (HM) release into soil solution: (a) without EDTA treatment; (b) after EDTA treatment. Different letters above bars indicate statistically significant differences at *p* < 0.05. (Modified from Jurkiewicz, A. et al., *Acta Biol. Cracov. Bot*., 46, 2004.)

This would suggest that mycorrhizal fungi increase the availability of metals to the plants but, at the same time, may decrease pollutant run-off into the ground water [\(Figure 13.3\).](#page-8-0)

Supplementing soil with sulphur can reduce the soil pH and the application of amendments and fertilizers can cause variations in the abundance of species [14] and modifications in the colonization of roots, thus increasing the amount of vesicles [15]. Also, the use of chelating chemicals, like EDTA, should be applied with much care: it mobilizes the toxic substances, but with the risk of leaching deeper into the soil profile.

In the general literature, not much attention has been paid to the fact that a large diversity in the effectiveness of phytoextraction might exist among different varieties of the same species, as shown by the previously mentioned study. These findings stress the necessity to screen a large number of cultivars in order to select the best ones for further use.

13.6 INFLUENCE OF SOIL BACTERIA ON MYCORRHIZA EFFICIENCY IN POLLUTED ENVIRONMENTS

Rhizosphere bacteria are known to improve mycorrhiza formation and activity by means of a number of so-called mycorrhizosphere activities, which benefit plant growth and health [126].

FIGURE 13.3 Heavy metal concentration in shoots of nonmycorrhizal and mycorrhizal *Plantago lanceolata* cultivated in rhizoboxes filled with zinc industrial wastes: 1: control plants (noninoculated); 2–6 — inoculated plants: 2: *Glomus clarum* isolated from zinc wastes; 3: *G. geosporum* from metal-polluted site; 4: *G. geosporum* from salt enriched site; 5: *G. claroideum* from zinc wastes; 6: *G. intraradices* from nonpolluted site. (From Orlowska, E. et al., *Polish Botanical Studies*, Polish Academy of Science [publisher] 2005.)

Therefore, it is to be expected that, with an appropriate selection of target bacteria, these could benefit the role of mycorrhiza in phytoremediation [127,128]. Selection procedures involve:

- Isolation of adapted bacteria from HM-contaminated soils
- Ecological compatibility with mycorrhizal fungi also adapted to HM contamination
- Functional compatibility of both types of microorganisms in terms of promoting phytoextraction and/or phytostabilization of metals from the polluted soil

Soil microbial diversity and activity are negatively affected by excessive concentration of HM [129]. However, indigenous bacterial populations must have adapted in a similar way to mycorrhizal fungi [14] and metal toxicity and evolved abilities that enable the bacteria to survive in polluted soils [127]. Adaptation of mycorrhizal fungi and associated bacteria to HM is considered a prerequisite for exploiting their potential role in phytoremediation [49].

The role of a tailored mycorrhizosphere in phytoremediation was investigated in a series of studies [130–134]. These studies consisted of:

- Isolation and characterization of microorganisms from a target HM contaminated site
- Development of several phytoremediation experiments
- Analysis of the mechanisms involved to account for the demonstrated phytoextraction and/or phytostabilization activities found

Under natural conditions, soil becomes contaminated with more than one metal; thus, it is difficult to determine which metals are responsible for the toxic effects observed [135]. Therefore, only long-term experiments using soils supplemented with a single metal salt can give the opportunity to study the individual toxic effects of each heavy metal on the beneficial microbes for a given time period [136]. In this context, a number of experiments are summarized in the present chapter, all of them using agricultural soil from Nagyhörcsök Experimental Station (Hungary). This soil was contaminated in 1991 with suspensions of 13 microelement salts applied separately. Each salt was applied at four levels $(0, 30, 90,$ and $270 \text{ mg kg}^{-1})$ as described by Biró et al. [136].

Indigenous mycorrhizal fungi and bacterial strains were isolated from this HM-polluted soil 10 years after contamination. They were tested for their influence on plant growth and on the functioning of native mycorrhizal fungi in the face of Cd, Pb, or Ni toxicity.

The most efficient bacterial isolates were identified by means of 16S rDNA sequence analysis and confirmed to belong to the genus *Brevibacillus*. Particularly, *B. brevis* was the most abundant species [130,132]. *Glomus mosseae* was present in all the HM-polluted soil samples, so it was the target mycorrhizal fungus used for phytoremediation inoculation experiments. *G. mosseae* and *Brevibacillus* sp. strains from nonpolluted environments were also used as reference inocula. *Trifolium repens* L. was used as test plant and inoculated with a suspension of *Rhizobium leguminosarum* bv *trifoli*, also an HM-tolerant strain.

In the Cd-contaminated soil [132,133], a high level functional compatibility between both types of autochthonous microorganisms was demonstrated; this resulted in a biomass increase of 545% (shoots) and 456% (roots) and in the N and P content compared to nonmycorrhizal plants. Coinoculation of both microorganisms increased root biomass and symbiotic structures (nodules and AM colonization) to a highest extent, which may be responsible for the beneficial effect observed. The results suggest that bacterial inoculation improved the mycorrhizal benefit in phytostabilization.

Dual inoculation of the Cd-adapted autochthonous *Brevibacillus* sp. and the AM fungus lowered the Cd concentration in *Trifolium* plants. This effect can be due to the ability of the bacteria to accumulate great amounts of Cd. In spite of that, the total Cd content accumulated in plant shoots was higher in dually inoculated plants. This indicates a phytoextraction activity resulting from such a dual inoculation. Further studies [134] demonstrated that the inoculated Cd-adapted bacteria increased dehydrogenase, phosphatase, and β-gluconase activities in the mycorrhizosphere, indicating an improvement of microbial activities concerning plant development in the polluted test soil.

With respect to Pb-spiked soil, experiments following the same methodological approaches [130] showed that *B. agri* at all Pb-spiking levels assayed consistently enhanced plant growth and nutrient accumulation in mycorrhizal plants, as well as nodule numbers and mycorrhizal colonization. This suggests a phytostabilization activity. Auxin production by the test bacteria can account for the beneficial role of these bacteria on mycorrhizal plant development [126]. Dual inoculation increased Pb concentration in plant shoots at the highest level of Pb applied. However, the total content of this metal in plants was consistently enhanced at all levels of Pb, showing that bacterial inoculation enhanced phytoextraction activities in plants inoculated with mycorrhizal fungi.

Dual inoculation of an indigenous Ni-adapted mycorrhizal strain of *G. mosseae* and a Niadapted bacterium (*Brevibacillus* sp.) isolated from Ni-contaminated soil was also assayed with *Trifolium* plants growing in Ni-polluted soil [131]. Dual inoculation increased total plant content of this metal at all levels of Ni assayed. This indicates that the tailored mycorrhizosphere carries out phytoextraction of Ni from polluted soils.

The mechanisms by which the bacterial isolates tested enhanced phytoremediation activities in mycorrhizal plants can be therefore summarized as follows:

- Improving rooting, mycorrhiza formation, and functioning
- Enhancing microbial activities in the mycorrhizosphere
- Accumulating metals, thus avoiding their transfer to the aquifers

In conclusion, the dual inoculation of suitable symbiotic and saprophytic rhizosphere microorganisms isolated from HM-polluted soils seems to play an important role in the development and HM tolerance by plants and in bioremediation of HM-contaminated soils.

13.7 MYCORRHIZA AS INDICATOR OF SOIL TOXICITY AND REMEDIATION RATE

Mycorrhizal fungi can also be useful as indicators of soil toxicity [71,137] and the effectiveness of remediation [76]. The toxicity of heavy metals and other pollutants (xenobiotics, PAH) has been monitored using AMF spore germination [13], mycorrhizal colonization of roots analyzed by PCR technique [138], and mycorrhizal infectivity [61]. Recently, the toxicity of zinc wastes of different ages and resulting from different extraction technologies has been compared using various AMF strains and species. The activity of the alkaline phosphatase [139], a vital staining of mycorrhizal colonization, has been found to provide a more sensitive test than the estimation of the total mycorrhizal development [57,62]. Similarly to other indicator organisms such as plants, earthworms, algae, and fish, the disadvantage is that all of them, including AMF, are not specific to pollutants and also react to soil properties (P and N content, pH, etc.).

The appropriate selection of the control soil seems to be problematic. The examples studied thus far prove that they are sensitive indicators of the changes that occur during phytoremediation or during spontaneous succession. Concerning a wider range of soils, the most useful is the germination test, especially that the technique is presently standardized and may rely on the fungal strains supplied commercially (Leyval, C., personal communication, 2004). Further approaches have been recently done to establish new methods considering a wide range of features, such as abundance of intraradical and extraradical mycelium; formation of vesicles; distribution of lipid droplets; etc. They seem to react sensitively to pollutants, at least in the case of the most widely used *Glomus mosseae* strain, BEG12.

Some AMF can be more sensitive to pollutants than plants [72], although some species, especially those originating from strongly polluted places, are well adapted to survival under extremely harsh conditions and their disappearance might be caused only by the lack of the symbiotic plants. The selection of an appropriate fungal strain and plant varieties is therefore of utmost importance [76]. Among the plant species analyzed thus far, English plantain (*Plantago lanceolata* L.), strongly colonized by arbuscular fungi, deserves special attention as an indicator used for bioassays. It occurs in diverse habitats and is resistant to a wide range of stress factors; it is also easy to obtain clones of one plant, which eliminates genetic variability among individuals in their response to toxic compounds [140–142].

13.8 CONCLUSIONS

Central and Eastern Europe are regions where large industrial wastes, deposits of various kinds of waste materials, and places polluted by insufficiently secured unused plant protection products as well as sites subjected to intense motorization and industrialization — are especially common. Despite the usually well-designed actions aiming at explaining the problem of pollution to the local community, one can still see the production of plants destined for human consumption on heavily polluted soils. Cheap and fast monitoring methods are needed here, followed by low-cost and effective phytoremediation techniques; mycorrhizal fungi should play the key role. It should be emphasized that research on this group of fungi should be conducted in a complex way and include other mycorhizosphere organisms with which the fungi interact [143].

The previously mentioned questions illustrate how broad and diverse are the possibilities to use natural phenomena in solving difficult problems of today's civilization. This will certainly stimulate the dynamic development of a range of scientific fields aimed at explaining the mechanisms of the mentioned phenomena and at optimizing their practical applications.

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REFERENCES

- 1. Adriano, D.C., Chlopecka, A. and Kaplan, D.I., Role of soil chemistry in soil remediation and ecosystem conservation, in *Soil Chemistry and Ecosystem Health*. Special Publication no. 52, Soil Science Society of America, Madison, WI, 1998, chap. 1.
- 2. Smith, S.E. and Read, D.J., *Mycorrhizal Symbiosis*, Academic Press, London, 1997, 605.
- 3. Linderman, R.G., Effects of mycorrhizas on plant tolerance to diseases, in *Arbuscular Mycorrhizas: Physiology and Function*, Kapulnik, Y. and Douds, D., Jr., Eds., Kluwer Academic Publishers, Dordrecht, The Netherlands, 2000, 345.
- 4. Turnau, K. and Haselwandter, K., Arbuscular mycorrhizal fungi, an essential component of soil microflora in ecosystem restoration, in *Mycorrhizal Technology in Agriculture. From Genes to Bioproducts*, Gianinazzi, S., Schepp, H., Barea, J.M. and Haselwandter, K., Eds., Birkhauser Verlag, Switzerland, 2002, 137.
- 5. Turnau, K. and Kottke, I., Fungal activity as determined by microscale methods with special emphasis on interactions with heavy metals, in *The Fungal Community*, Dighton, J., Ed., Marcel Dekker, 2005, Chapter 14.
- 6. Azcón–Aguilar, C. and Barea, J.M., Arbuscular mycorrhizas and biological control of soil-borne plant pathogens. An overview of the mechanisms involved, *Mycorrhiza*, 6, 457, 1996.
- 7. Barea, J.M., Mycorrhiza/bacteria interactions on plant growth promotion, in *Plant Growth-Promoting Rhizobacteria, Present Status and Future Prospects*, Ogoshi, A., Kobayashi, L., Homma, Y., Kodama, F., Kondon, N. and Akino, S., Eds., OECD, Paris, 1997,150.
- 8. Barea, J.M., Rhizosphere and mycorrhiza of field crops, in *Biological Resource Management*: *Connecting Science and Policy*, Toutant, J.P., Balazs, E., Galante, E., Lynch, J.M., Schepers, J.S., Werner, D. and Werry, P.A., Eds., OECD, INRA edition and Springer, 2000, 110.
- 9. Miller, R.M. and Jastrow, J.D., Mycorrhizal fungi influence soil structure, in *Arbuscular Mycorrhizas: Physiology and Function*, Kapulnik, Y. and Douds, D.D., Eds., Kluwer Academic Publishers, Netherlands, 2000, 3.
- 10. Schussler, A., Schwarzott, D. and Walker, C., A new fungal phylum, the Glomeromycota: phylogeny and evolution, *Mycol. Res*., 105, 1413, 2001.
- 11. Remy, W., Taylor, T.N., Haas, H. and Kerp, H., Four-hundred-million-year-old vesicular-arbuscular mycorrhizae, *Proc. Natl. Acad. Sci*., 91, 11841, 1994.
- 12. Simon, L., Bousquet, J., Lévesque, R.C. and Lalonde, M., Origin and diversification of endomycorrhizal fungi and coincidence with vascular land plants, *Nature*, 363, 67, 1993.
- 13. Weissenhorn, I. and Leyval, C., Spore germination of arbuscular mycorrhizal fungi in soils differing in heavy metal content and other parameters, *Eur. J. Soil Biol.*, 32, 165, 1996.
- 14. Del Val, C., Barea, J.M. and Azcon–Aguilar, C., Diversity of arbuscular mycorrhizal fungus populations in heavy metal contaminated soils, *Appl. Environ. Microb*., 65, 718, 1999.
- 15. Pawlowska, T.E., Chaney, R.L., Chin, M. and Charvat, I., Effects of metal phytoextraction practices on the indigenous community of arbuscular mycorrhizal fungi at a metal-contaminated landfill, *Appl. Environ. Microbiol*., 66, 2526, 2000.
- 16. Blaylock, M.J., Zakharova, O., Salt, D.E. and Raskin, I., Increasing heavy metal uptake through soil amendments. The key to effective phytoremediation, in *Agronomy Abstracts*, ASA, Madison, WI, 1995, 218.
- 17. Salt, D.E., Blaylock, N., Kumar, N., Dushenkov, V., Ensley, B.D., Chet, I. and Raskin, I., Phytoremediation: a novel strategy for the removal of toxic metals from the environment using plants, *Biotechnology*, 13, 468, 1995.
- 18. Chlopecka, A. and Adriano, D.C., Mimicked *in situ* stabilization of metals in a cropped soil. *Environ. Sci. Technol*., 30, 3294, 1996.
- 19. Baker, A.J.M., Reeves, R.D. and Hajar, A.S.M., Heavy metal accumulation and tolerance in British populations of the metallophyte *Thlaspi caerulescens* J. and C. Presl. (Brassicaceae), *New Phytol*., 127, 61, 1994.
- 20. Baker, A.J.M. and Brooks, R.R., Terrestrial higher plants which hyperaccumulate metallic elements: a review of their distribution, ecology and phytochemistry, *Biorecovery*, 1, 81, 1989.
- 21. Levebre, K.K., Miki, B.L. and Laliberte, J.F., Mammalian metallothionein functions in plants, *Biotechnology*, 5, 1053, 1987.
- 22. Misra, S. and Gedamu, L., Heavy metal tolerant *Brassica napus* L. and *Nicotiana tabacum* L. plants, *Theor. Appl. Genet*., 78, 161, 1989.
- 23. Maiti, I.B., Wagner, G.J. and Hunt, A.G., Light inducible and tissue specific expression of a chimeric mouse metallothionein cDNA gene in tobacco, *Plant Sci.*, 76, 99, 1991.
- 24. Ernst, W.H.O., Bioavailability of heavy metals and decontamination of soils by plants, *Appl. Geochem*., 11, 163, 1996.
- 25. Rao Gadde, R. and Laitinen, H.A., Studies of heavy metal adsorption by hydrous iron and manganese oxides, *Anal. Chem*., 46, 2022, 1974.
- 26. Krishnamurti, G.S.R., Cieslinski, G., Huang, P.M., Van Rees, K.C.J., Kinetics of cadmium release from soils as influenced by organic acids: Implication in cadmium availability, *J. Environ. Qual*., 26, 271, 1997.
- 27. Lodenius, M. and Autio, S., Effects of acidification on the mobilization of cadmium and mercury from soils, *Arch. Environ. Con. Tox*., 18, 261, 1989.
- 28. Bloomfield, C., The translocation of metals in soils, in *The Chemistry of Soil Processes*, Greenland, D.J. and Hayes, M.H.B. Eds., John Wiley & Sons Ltd, Chichester, U.K., 1981, 50.
- 29. Fein, J.B., Daughney, C.J., Yee, N. and Davis, T.A., A chemical equilibrium model for metal absorption onto bacterial surfaces, *Geochem. Cosmochim. Acta*, 61, 3319, 1997.
- 30. Losi, M.E., Amrhein, C. and Frankenberger, W.T., Bioremediation of chromate contaminated groundwater by reduction and precipitation in surface soils, *J. Environ. Qual*., 23, 1141, 1994.
- 31. Kumar, P., Duschenkov, V., Motto, H. and Raskin, I., Phytoextraction: the use of plants to remove heavy metals from soils, *Environ. Sci. Technol*., 29, 1232, 1995.
- 32. Brooks, R.R., Plant hyperaccumulators of metals and their role in mineral exploration, archaeology, and land reclamation, in *Remediation of Metal-Contaminated Soils*, Iskandar, I.K. and Adriano, D.C., Eds., Science Reviews, Northwood, England, 1997, 123.
- 33. Gleba, D., Borisjuk, N.V., Borisjuk, L.G., Kneer, R., Poulev, A., Skarzhinskaya, M., Dushenkov, S., Logendra, S., Gleba, Y.Y. and Raskin, I., Use of plant roots for phytoremediation and molecular farming, *P. Natl. Acad. Sci. USA*, 96, 5973, 1999.
- 34. Allen, M.F., Sexton, J.C., Moore, T.S. and Christensen, M., Comparative water relations and photosynthesis of mycorrhizal and nonmycorrhizal *Bouteloua gracilis* (HBK) Lag ex Steud., *New Phytol*., 88, 683, 1981.
- 35. Allen, M.F., Influence of vesicular–arbuscular mycorrhizae on water movement through *Bouteloua gracilis* (HBK) Lag ex Steud., *New Phytol.*, 91, 191, 1982.
- 36. Nelsen, C. and Safir, G.R., The water relations of well watered, mycorrhizal and nonmycorrhizal onion plants, *J. Am. Soc. Hortic. Sci*., 107, 271, 1982.
- 37. Huang, R.S., Smith, W.K. and Yost, R.S., Influence of vesicular-arbuscular mycorrhiza on growth, water relations, and leaf orientation in *Leucaena leucocephala* (Lam.) De Wit., *New Phytol*., 99, 229, 1985.
- 38. Koide, R., The effect of mycorrhizal infection and phosphorus status on sunflower hydraulic and stomata properties, *J. Exp. Bot*., 36, 1087, 1985.
- 39. Fitter, A.H., Water relations of red clover, *Trifolium pratense* L., as affected by VA mycorrhizal infection and phosphorus supply before and during drought, *J. Exp. Bot*., 39, 595, 1988.
- 40. Berta, G., Fusconi, A., Trotta, A. and Scannerini, S., Morphogenic modifications induce by the mycorrhizal fungus *Glomus* strain E3 in the root system of *Allium porrum* L., *New Phytol*., 114, 207, 1990.
- 41. Kothari, S.K., Marschner, H. and George, E., Effect of VA mycorrhizal fungi and rhizosphere microorganisms on root and shoot morphology, growth and water relations in maize, *New Phytol*., 116, 303, 1990.
- 42. Baas, R. and Kuiper, D., Effects of vesicular–arbuscular mycorrhizal infection and phosphate on *Plantago major* ssp. *pleiosperma* in relation to internal cytokinin concentration, *Physiol. Plantarum,* 76, 211, 1989.
- 43. Danneberg, G., Latus, C., Zimmer, W., Hundeshagen, B., Schneider–Poetch, H.J. and Bothe, H., Influence of vesicular–arbuscular mycorrhizae on phytohormone balances in maize (*Zea mays* L.), *J. Plant Physiol*., 141, 33, 1992.
- 44. Torelli, A., Trotta, A., Acerbi, L., Arcidiacono, G., Berta, G. and Branca, C., IAA and ZR content in leek (*Allium porrum* L.) as influenced by P nutrition and arbuscular mycorrhizae, in relation to plant development, *Plant Soil*, 226, 29, 2000.
- 45. Burkert, B. and Robson, A.D., 65Zn uptake in subterranean clover (*Trifolium subterraneum* L.) by three vesicular–arbuscular mycorrhizal fungi in a root-free sandy soil, *Soil Biol. Biochem.*, 26, 1117, 1994.
- 46. Joner, E.J. and Leyval, C., Uptake of 109Cd by roots and hyphae of *Glomus mossae/Trifolium subterraneum* mycorrhiza from soil amended with high and low concentration of cadmium, *New Phytol*., 135, 353, 1997.
- 47. Kaldorf, M., Kuhn, A.J., Schroeder, W.H., Hildebrandt, U. and Bothe, H., Selective element deposit in maize colonized by heavy metal tolerance conferring arbuscular mycorrhizal fungus, *J. Plant Physiol*., 154, 718, 1999.
- 48. Shetty, K.G., Banks, M.K., Hetrick, B.A. and Schwab, A.P., Biological characterization of a southeast Kansas mining site, *Water Air Soil Pollut*., 78, 169, 1994.
- 49. Shetty, K.G., Hetrick, B.A.D., Figge, D.A.H. and Schwab, A.P., Effects of mycorrhizae and other soil microbes on revegetation of heavy metal contaminated mine spoil, *Environ. Pollut.*, 86, 181, 1994.
- 50. Turnau, K., Heavy metal uptake and arbuscular mycorrhiza development of *Euphorbia cyparissias* on zinc wastes in South Poland, *Acta Soc. Bot. Pol.*, 67, 105, 1998.
- 51. El-Kherbawy, M., Angle, J.S., Heggo, A. and Chaney, R.L., Soil pH, rhizobia and vesicular-arbuscular mycorrhizae inoculation effects on growth and heavy metal uptake of alfalfa (*Medicago sativa* L.), *Biol. Fert. Soils*, 8, 61, 1989.
- 52. Meharg, A.A. and Cairney, J.W.G., Ectomycorrhizas extending the capabilities of rhizosphere remediation? *Soil Biol. Biochem*., 32, 1475, 2000.
- 53. Leyval, C., Turnau, K. and Haselwandter, K., Effect of heavy metal pollution on mycorrhizal colonization and function: physiological, ecological and applied aspects, *Mycorrhiza*, 7, 139, 1997.
- 54. Lanfranco, L., Bolchi, A., Cesale Ros, E., Ottonello, S. and Bonfante, P., Differential expression of a metallothionein gene during the presymbiotic vs. the symbiotic phase of an arbuscular mycorrhizal fungus, *Plant Physiol*., 130, 58, 2002.
- 55. Joner, E.J., Briones, R. and Leyval, C., Metal-binding capacity of arbuscular mycorrhizal mycelium, *Plant Soil*, 226, 227, 2000.
- 56. Hildebrandt, U., Kaldorf, M. and Bothe, H., The zinc violet and its colonization by arbuscular mycorrhizal fungi, *J. Pl. Physiol*., 154, 709, 1999.
- 57. Orlowska, E., Jurkiewicz,A., Anielska, T., Godzik,B. and Turnau, K., Influence of different arbuscular mycorrhizal fungal (AMF) strains on heavy metal uptake by *Plantago lanceolata* L., *Polish Botanical Studies*, Polish Academy of Science, publisher, 19, 65, 2006.
- 58. Tonin, C., Vandenkoornhuyse, P., Joner, E.J., Straczek, J. and Leyval, C., Assessment of arbuscular mycorrhizal fungi diversity in the rhizosphere of *Viola calaminaria* and effect of these fungi on heavy metal uptake by clover, *Mycorrhiza*, 10, 161, 2001.
- 59. Rivera–Becerril, F., Calantzis, C., Turnau, K., Caussanel, J-P., Belimov, A.A., Gianinazzi, S., Strasser, R.J. and Gianinazzi–Pearson, V.J., Cadmium accumulation and buffering of cadmium-induced stress by arbuscular mycorrhiza in three *Pisum sativum* L. genotypes, *Exp. Bot*., 53(371), 1177, 2002.
- 60. Galli, U., Schuepp, H. and Brunold, C., Heavy metal binding by mycorrhizal fungi, *Physiol. Pl*. 92, 364, 1994.
- 61. Leyval, C., Singh, B.R. and Joner, E.J., Occurrence and infectivity of arbuscular mycorrhizal fungi in some Norwegian soils influenced by heavy metals and soil properties, *Water Air Soil Pollut*., 83, 203, 1995.
- 62. Orlowska, E., Ryszka, P., Jurkiewicz, A. and Turnau, K., Effectiveness of arbuscular mycorrhizal fungal (AMF) strains in colonization of plants involved in phytostabilisation of zinc wastes, *Geoderma*, 2005, in press.
- 63. Turnau, K., Ryszka, P., Van Tuinen, D. and Gianinazzi-Pearson, V., Identification of arbuscular mycorrhizal fungi in soils and roots of plants colonizing zinc wastes in Southern Poland, *Mycorrhiza*, 10, 169, 2001.
- 64. Van Tuinen, D., Jacquot, E., Zhao, B., Gallotte, A. and Gianinazzi-Pearson, V., Characterization of root colonization profiles by a microcosm community of arbuscular mycorrhizal fungi using 25S rDNA-targeted nested PCR, *Mol. Ecol*., 7, 879, 1998.
- 65. Jacquot–Plumey, E., van Tuinen, D., Chatagnier, O., Gianinazzi, S. and Gianinazzi–Pearson, V., 25S rDNA-based molecular monitoring of glomalean fungi in sewage sludge-treated field plots, *Environ. Microbiol*., 3, 525, 2001.
- 66. Antosiewicz, D.M., Adaptation of plants to an environment polluted with heavy metals, *Acta Soc. Bot. Pol.*, 61, 281, 1992.
- 67. Koide, R.T. and Schreiner, R.P., Regulation of the vesicular-arbuscular mycorrhizal symbiosis, *Annu. Rev. Pl. Physiol. Pl. Mol. Biol*., 43, 557, 1992.
- 68. Daft, M.J. and Nicolson, T.H., Arbuscular mycorrhizas in plants colonizing coal wastes in Scotland, *New Phytol*., 73, 1129, 1974.
- 69. Daft, M.J., Hacskaylo, E. and Nicolson, T.H., Arbuscular mycorrhizas in plants colonizing coal spoils in Scotland and Pennsylvania, in *Endomycorrhizas*, Sanders, F.E., Mosse, B. and Tinker, P.B., Eds., Academic Press, London, 1975, 561.
- 70. Pawlowska, T.E., Baszkowski, J. and Rühling, A., The mycorrhizal status of plants colonizing a calamine spoil mound in southern Poland, *Mycorrhiza*, 6, 499, 1996.
- 71. Gucwa–Przepióra, E. and Turnau, K., Arbuscular mycorrhiza and plant succession in the zinc smelter spoil heap in Katowice–Wenowiec, *Acta Soc. Bot. Pol*., 70/2, 153, 2001.
- 72. Weissenhorn, I. and Leyval, C., Root colonization of maize by a Cd-sensitive and a Cd-tolerant *Glomus mosseae* and cadmium uptake in sand culture, *Plant Soil*, 175, 233, 1995.
- 73. Noyd, R.K., Pfleger, F.L. and Norland, M.R., Field responses to added organic matter, arbuscular mycorrhizal fungi, and fertilizer in reclamation of turbonite iron ore tailing, *Plant Soil*, 179, 89, 1996.
- 74. Khan, A.G., Kuek, C., Chaudhry, T.M., Khoo, C.S. and Hayes, W.J., Role of plants, mycorrhizae and phytochelators in heavy metal contaminated land remediation, *Chemosphere*, 41, 197, 2000.
- 75. Vosatka, M., Investigation of VAM in *Sorbus aucuparia* and *Acer pseudoplatanus* stands on airpolluted localities and mine spoils in North Bohemia, *Agr. Ecosyst. Environ*., 29, 443, 1989.
- 76. Orlowska, E., Zubek, Sz., Jurkiewicz, A., Szarek–Sukaszewska, G. and Turnau, K., Influence of restoration on arbuscular mycorrhiza of *Biscutella laevigata* L. (Brassicaceae) and *Plantago lanceolata* L. (Plantaginaceae) from calamine spoil mounds, *Mycorrhiza*, 12, 153, 2002.
- 77. Haselwandter, K. and Bowen, G.D., Mycorrhizal relations in trees for agroforestry and land rehabilitation, *Forest Ecol. Manage*., 81, 1, 1996.
- 78. Hartley–Whitaker, J., Cairney, J.W.G. and Meharg, A.A., Sensitivity to Cd or Zn of host and symbiont of ectomycorrhizal *Pinus sylvestris* L. (Scots pine) seedlings, *Plant Soil*, 218, 31, 2000.
- 79. Blaudez, D., Jacob, C., Turnau, K., Colpaert, J.V., Ahonen–Jonnarth, U., Finlay, R., Botton, B. and Chalot, M., Differencial responses of ectomycorrhizal fungi to heavy metals *in vitro*, *Mycol. Res*., 104, 1366, 2000.
- 80. Colpaert, J.V., Vandenkoornhuyse, P., Adriansen, K. and Vangronsveld, J., Genetic variation and heavy metal tolerance in the ectomycorrhizal basidiomycete *Suillus luteus*, *New Phytol*., 147, 367, 2000.
- 81. Turnau, K., Kottke, I., Dexheimer, J. and Botton, B., Element distribution in *Pisolithus tinctorius* mycelium treated with cadmium dust, *Ann. Bot*., 74, 137, 1994.
- 82. Galli, U., Meier, M. and Brunold, C., Effects of cadmium on nonmycorrhizal and mycorrhizal Norway spruce seedlings (*Picea abies* (L.) Karst.) and its ectomycorrhizal fungus *Laccaria laccata* (Scop. ex Fr.) Bk. and Br.: sulphate reduction, thiols and distribution of the heavy metals, *New Phytol*., 125, 837, 1993.
- 83. Tam, P.C.F., Heavy metal tolerance by ectomycorrhizal fungi and metal amelioration by *Pisolithus tinctorius*, *Mycorrhiza*, 5, 181, 1995.
- 84. Turnau, K., Kottke, I. and Dexheimer, J., Toxic element filtering in *Rhizopogon roseolus/Pinus sylvestris* mycorrhizas collected from calamine dumps, *Mycol. Res*., 100/1, 16, 1996.
- 85. Turnau, K., Przybylowicz, W.J. and Mesjasz–Przybylowicz, J., Heavy metal distribution in *Suillus luteus* mycorrhizas *—* as revealed by micro-PIXE analysis, *J. Nucl. Instruments*, 181, 649, 2001.
- 86. Wilson, N.G. and Bradley, G., Enhanced degradation of petrol (Slovene diesel) in an aqueous system by immobilized *Pseudomonas fluorescens*, *J. Appl. Bacteriol*., 80(1), 99, 1996.
- 87. Bezalel, L., Hadar, Y. and Cerniglia, C.E., Enzymatic mechanisms involved in phenanthrene degradation by the white rot fungus *Pleurotus ostreatus*, *Appl. Environ. Microbiol*., 63, 2495, 1997.
- 88. Schützendübel, A., Majcherczyk, A., Johannes, C. and Huttermann, A., Degradation of fluorene, anthracene, phenanthrene, fluoranthene, and pyrene lacks connection to the production of extracellular enzymes by *Pleurotus ostreatus* and *Bjerkandera adusta*, *Int. Biodeter. Biodegr*., 43(3), 93, 1999.
- 89. Schwab, A.P. and Banks, M.K., Biologically mediated dissipation of polyaromatic hydrocarbons in the root zone, in *Bioremediation through Rhizosphere Technology*, Anderson, T.A. and Coats, J.R., Eds., American Chemical Society, Washington D.C., 1994, 132.
- 90. Reilley, K.A., Banks, M.K. and Schwab, A.P., Dissipation of polycyclic aromatichydrocarbons in the rhizosphere, *J. Environ. Qual*., 25, 212, 1996.
- 91. Shann, J.R. and Boyle, J.J., Influence of plant species on *in situ* rhizosphere degradation, in *Bioremediation through Rhizosphere Technology*, Anderson, T.A. and Coats, J.R., Eds., American Chemical Society, Washington D.C., 1994, 70.
- 92. Leyval, C., Joner, E.J., del Val, C. and Haselwandter, K., Potential of arbuscular mycorrhizal fungi for bioremediation, in *Mycorrhizal Technology in Agriculture. From Genes to Bioproducts,* Gianinazzi, S., Schuepp, H., Barea, J.M. and Haselwandter K., Eds., Birkhauser Verlag, Switzerland, 2002, 175.
- 93. Leyval, C. and Binet, P., Effect of polyaromatic hydrocarbons in soil on arbuscular mycorrhizal plants, *J. Environ. Qual*., 27, 402, 1998.
- 94. Joner, E.J. and Leyval, C., Influence of arbuscular mycorrhiza on clover and ryegrass grown together in a soil spiked with polycyclic aromatic hydrocarbons, *Mycorrhiza*, 10, 155, 2001.
- 95. Sanchez–Diaz, M. and Honrubia, M., Water relations and alleviation of drought stress in mycorrhizal plants, in *Impact of Arbuscular Mycorrhizas on Sustainable Agriculture and Natural Ecosystems*, Gianinazzi, S. and Schüepp, H., Eds., Birkhäuser, Basel, 1994, 167.
- 96. Salzer, P., Corbere, H. and Boller, T., Hydrogen peroxide accumulation in *Medicago truncatula* roots colonized by the arbuscular mycorrhiza-forming fungus *Glomus intraradices*, *Planta,* 208, 319, 1999.
- 97. Barr, D.P. and Aust, S.D., Mechanisms white rot fungi use to degrade pollutants, *Environ. Sci. Technol.*, 28, 79, 1994.
- 98. Donnelly, P.K. and Fletcher, J.S., Potential use of mycorrhizal fungi as bioremediation agents, in *Bioremediation through Rhizosphere Technology*, Anderson, T.A. and Coats, J. R., Eds., American Chemical Society, Washington D.C., 1994, 93.
- 99. Gilbert, E.S. and Crowley, D.E., Plant compounds that induce polychlorinated biphenyl biodegradation by *Arthrobacter* sp. strain B1B, *Appl. Environ. Microbiol*., 63, 1933, 1997.
- 100. Green, N.A., Meharg, A.A., Till, C., Troke, J. and Nicholson, J.K., Degradation of 4-fluorobiphenyl by mycorrhizal fungi as determined by 19C radiolabelling analysis, *Appl. Environ. Microbiol*., 65, 4021, 1999.
- 101. Dias, S.M., Tratamento de efluentes em zonas humidas construidas ou leitos de macrofitas, *Bol. Biotechnol*., 60, 14, 1998.
- 102. Trautmann, N., Martin, J.H., Porter, K.S. and Hawk, K.C., Use of artificial wetlands treatment of municipal solid waste landfill leachate, in *Constructed Wetlands for Waste Water Treatment: Municipal, Industrial and Agricultural*, Hammer, D.A., Ed., Lewis, Chelsea, MI, 1989, 245.
- 103. Maehlum, T., Treatment of landfill leachate in on-site lagoons and constructed wetlands, *Water Sci. Technol*., 32(3), 129, 1995.
- 104. Oliveira, R.S., Dodd, J.C. and Castro, P.M.L., The mycorrhizal status of *Phragmites australis* in several polluted soils and sediments of an industrialized region of Northern Portugal, *Mycorrhiza,* 10(5), 241, 2001.
- 105. Baker, A.J.M., McGrath, S.P., Reeves, R.D. and Smith, J.A.C., Metal hyperaccumulator plants: a review of the ecology and physiology of a biological resource for phytoremediation of metal polluted soils, in *Phytoremediation of Contaminated Soil and Water*, Terry, N. and Banuelos, G., Eds., CRC Press, Boca Raton, FL, 2000, 85.
- 106. Salt, D.E., Smith, R.D. and Raskin, I., Phytoremediation, *Annu. Rev. Plant Phys*., 49, 643, 1998.
- 107. Nicks, L. and Chambers, M.F., A pioneering study of the potential of phytomining for nickel, in *Plants That Hyperaccumulate Heavy Metals — Their Role in Phytoremediation*, *Microbiology*, *Archeology*, *Mineral Exploration and Phytomining*, Brooks, R.R., Ed., CAB International, Wallingford, MA, 1998, 313.
- 108. Brooks, R.R. and Robinson, B.H., The potential use of hyperaccumulators and other plants for phytomining, in *Plants That Hyperaccumulate Heavy Metals — Their Role in Phytoremediation*, *Microbiology*, *Archeology*, *Mineral Exploration and Phytomining*, Brooks, R.R. Ed., CAB International, Cambridge, MA, 1998, 327.
- 109. Chaney, R.L., Mallik, M., Li, Y.M., Brown, S.L. and Brewer, E.P., Phytoremediation of soil metals, *Curr. Opin. Biotech*., 8, 279, 1997.
- 110. Chaney, R.L., Lee, Y.M., Brown, S.L., Homer, F.A., Malik, M., Angle, J.S., Baker, A.J.M., Reeves, R.D. and Chin, M., Improving metal hyperaccumulator wild plants to develop commercial phytoextraction systems: approaches and progress, in *Phytoremediation of Contaminated Soil and Water*, Terry, N. and Banuelos, G., Eds., CRC Press, Boca Raton, FL, 2000, 129.
- 111. Goncalves, S.C., Goncalves, M.T., Freitas, H. and Martins Loucao, M.A., Mycorrhizae in a Portuguese serpentine community, in *The Ecology of Ultramafic and Metalliferous Areas,* Jaffre, T., Reeves, R.D. and Becquer, T., Eds., *Proc. Second Int.Conf. Serpentine Ecol*. in Noumea (1995), 1997, 87.
- 112. Hopkins, N.A., Mycorrhizae in a California serpentine grassland community, *Can. J. Bot*., 65, 484, 1987.
- 113. Augustyniak, M., Mesjasz-Przybylowicz, J., Nakonieczny, M., Dybowska, M., Przybylowicz, W. and Migula, P., Food relations between *Chrysolina pardalina* and *Berkheya coddii*, a nickel hyperaccumulator from South African ultramafic outcrops, *Fresen. Environ. Bull.*, 11(2), 85, 2002.
- 114. Regvar, M., Vogel, K., Irgel, N., Wraber, T., Hildebrandt, U., Wilde, P. and Bothe, H., Colonization of pennycresses (*Thlaspi* spp.) of the *Brassicaceae* by arbuscular mycorrhizal fungi, *J. Plant Physiol*., 160(6), 615, 2003.
- 115. Huang, J.W. and Cunningham, S.D., Lead phytoextraction: species variation in lead uptake and translocation, *New Phytol*., 134, 75, 1996.
- 116. Huang, J.W., Chen, J., Berti, W.R. and Cunningham, S.D., Phytoremediation of lead-contaminated soils: role of synthetic chelates in lead phytoextraction, *Environ. Sci. Technol*., 31, 800, 1997.
- 117. Rensing, C., Sun, Y., Mitra, B. and Rosen, B.P., Pb(II)-translocating P-type ATPases, *J. Biol. Chem*., 273, 32614, 1998.
- 118. Karenlampi, S., Schat, H., Vangronsveld, J., Verkleij, J.A.C., Van Der Lelie, D., Mergeay, M. and Tervahauta, A.I., Genetic engineering in the improvement of plants for phytoremediation of metal polluted soils, *Environ. Pollut*., 107, 225, 2000.
- 119. Riba, G. and Chupeau, Y., Genetically modified plants, *Cell. Mol. Biol*., 47, 1319, 2001.
- 120. Brewer, E.P., Sauders, J.A., Angle, J.S., Chaney, R.L. and Mcintosh, M.S., Somatic hybridization between the zinc accumulator *Thlaspi caerulescens* and *Brassica napus*, *Theor. Appl. Genet*., 99, 761, 1999.
- 121. Krämer, U. and Chardonnens, A.N., The use of transgenic plants in the bioremediation of soils contaminated with trace elements, *Appl. Microbiol. Biotechnol*., 55, 661, 2001.
- 122. Harrier, L., Millam, S., Biolistic transformation of arbuscular mycorrhizal fungi: progress and perspectives, *Mol. Biotechnol.*, 18, 25, 2001.
- 123. Koch, A.M., Kuhn, G., Fontanillas, P., Fumagalli, L., Goudet, I. and Sanders, I.R., High genetic variability and low local diversity in a population of arbuscular mycorrhizal fungi, *Proc. Natl. Acad. Sci*., 101, 2369, 2004.
- 124. Blaylock, M.J., Salt, D.E., Dushenkov, S., Zakharova, O., Gussman, C., Kapulnik, Y., Ensley, B.D. and Raskin, I., Enhanced accumulation of Pb in Indian mustard by soil-applied chelating agents, *Environ. Sci. Technol*., 31, 860, 1997.
- 125. Jurkiewicz, A., Orlowska, E., Anielska, T., Godzik, B. and Turnau, K., The influence of mycorrhiza and EDTA application on heavy metal uptake by different maize varieties, *Acta Biol. Cracov. Bot*., 46, 7, 2004.
- 126. Barea, J.M., Gryndler, M., Lemanceau, Ph., Schüepp, H. and Azcón, R., The rhizosphere of mycorrhizal plants, in *Mycorrhiza Technology in Agriculture. From Genes to Bioproducts*, Gianinazzi, S., Schüepp, H., Barea, J.M. and Haselwandter, K., Eds., Birkhäuser Verlag, Basel, Switzerland, 2002, 1.
- 127. Biró, B., Bayoumi, H.E.A.F., Balázsy, S. and Kecskés, M., Metal sensitivity of some symbiotic N2 fixing bacteria and *Pseudomonas* strains, *Acta Biol. Hung*., 46, 9, 1995.
- 128. Takács, T., Biró, B., Vörös., I., Arbuscular mycorrhizal effect on heavy metal uptake of ryegrass (*Lolium perenne* L.) in pot culture with polluted soil, in *Plant Nutrition Food Security and Sustainability of Agro-Ecosystems*, Horst, W.J. Ed., Kluwer Academic Publishers, The Netherlands, 2001, 480.
- 129. Giller, K., Witter, E. and McGrath, S., Toxicity of heavy metals to microorganisms and microbial processes in agricultural soils: a review, *Soil. Biol. Biochem*., 30, 1389, 1998.
- 130. Vivas, A., Azcón, R., Biró, B., Barea, J.M. and Ruiz-Lozano, J.M., Influence of bacterial strains isolated from lead-polluted soil and their interactions with arbuscular mycorrhizae on the growth of *Trifolium pratense* L. under lead toxicity, *Can. J. Microbiol*., 49, 577, 2003.
- 131. Vivas, A., Biró, B., Anton, A., Vörös, I., Barea, J.M. and Azcón, R., Possibility of phytoremediation by co-inoculated Ni-tolerant mycorrhiza-bacterium strains, in *Trace Elements in the Food Chain*, Simon, L. and Szilágyi György, M., Eds., Bessenyei Publisher, Nyíregyháza, Hungary, 2003, 76.
- 132. Vivas, A., Vörös, I., Biró, B., Barea, J.M., Ruiz-Lozano, J.M. and Azcón, R., Beneficial effects of indigenous Cd-tolerant and Cd-sensitive *Glomus mosseae* associated with a Cd-adapted strain of *Brevibacillus* sp. in improving plant tolerance to Cd contamination, *Appl. Soil Ecol*., 24, 177, 2003.
- 133. Vivas, A., Vörös, I., Biró, B., Campos, E., Barea, J.M. and Azcón, R., Symbiotic efficiency of autochthonous arbuscular mycorrhizal fungus (*G. mosseae*) and *Brevibacillus* sp. isolated from cadmium polluted soil under increasing cadmium levels, *Environ. Pollut*., 126, 179, 2003.
- 134. Vivas, A., Barea, J.M. and Azcón, R., Interactive effects of *Brevibacillus brevis* and *Glomus mosseae*, both from a Cd contaminated soil, on plant growth, physiological mycorrhizal characteristic and soil enzymatic activities in a Cd- spiked soil, *Environ. Pollut*. 134, 275, 2004.
- 135. Chaudri, A.M., McGrath, S.P. and Giller, K.E., Survival of the indigenous population of *Rhizobium leguminosarum* bv. *trifolii* in soil spiked with Cd, Zn, Cu and Ni salts, *Soil Biol. Biochem*., 24, 625, 1992.
- 136. Biró, B., Köves–Péchy, K., Vörös, I. and Kádár, I., Toxicity of some field applied heavy metal salts to the rhizobial and fungal microsymbionts of alfalfa and red clover, *Agrokém. Talajtan*, 47, 265, 1998.
- 137. Weissenhorn, I., Leyval, C. and Berthelin, J., Cd-tolerant arbuscular mycorrhizal (AM) fungi from heavy metal polluted soils, *Plant Soil*, 157, 247, 1993.
- 138. Jacquot–Plumey, E., van Tuinen, D., Gianinazzi, S., Gianinazzi–Pearson, V., Monitoring species of arbuscular mycorrhizal fungi in planta and in soil by nested PCR: application to the study of the impact of sewage sludge, *Plant Soil*, 226,179, 2000.
- 139. Van Aarle, I.M., Olsson, P.A., and Söderström, B., Microscopic detection of phosphatase activity of saprophytic and arbuscular mycorrhizal fungi using a fluorogenic substrate, *Mycologia*, 93(1), 17, 2001.
- 140. Wu, L. and Antonovics, J., Experimental ecological genetics in *Plantago*. I. Induction of roots and shoots on leaves for large scale vegetative propagation and metal tolerance testing in *P. lanceolata*, *New Phytol*., 75, 277, 1975.
- 141. Baroni, F., Boscagli, A., Protano, G. and Riccobono, F., Antimony accumulation in *Achillea ageratum*, *Plantago lanceolata* and *Silene vulgaris* growing in an old Sb-mining area, *Environ. Pollut.*, 109, 347, 2000.
- 142. Bakker, M.I., Vorenhout, M., Sijm, D.T.H.M. and Kollofel, C., Dry deposition of atmospheric polycyclic hydrocarbons in three *Plantago* species, *Environ. Toxicol. Chem.*, 18, 2289, 1999.
- 143. Jeffries, P., Gianinazzi, S., Perotto, S., Turnau, K. and Barea, J.M., The contribution of arbuscular mycorrhizal fungi in sustainable maintenance of plant health and soil fertility, *Biol. Fertil. Soils*, 37, 1, 2003.